# Water-Soluble Naphthalene Diimides as Singlet Oxygen Sensitizers

Filippo Doria,<sup>†</sup> Ilse Manet,<sup>\*,‡</sup> Vincenzo Grande,<sup>†</sup> Sandra Monti,<sup>‡</sup> and Mauro Freccero<sup>\*,†</sup>

<sup>†</sup>Dipartimento di Chimica, Università di Pavia, V. le Taramelli 10, 27100 Pavia, Italy

<sup>‡</sup>Istituto per la Sintesi Organica e la Fotoreattività, Consiglio Nazionale delle Ricerche, via Gobetti 101, 40129 Bologna, Italy

**Supporting Information** 

**ABSTRACT:** Bromo- and/or alkylamino-substituted and hydrosoluble naphthalene diimides (NDIs) were synthesized to study their multimodal photophysical and photochemical properties. Bromine-containing NDIs (i.e., **11**) behaved as both singlet oxygen ( $^{1}O_{2}$ ) photosensitizers and fluorescent molecules upon irradiation at 532 nm. Among the NDIs not containing Br, only **12** exhibited photophysical properties similar to those of Br-NDIs, by irradiation above 610 nm, suggesting that for these NDIs both singlet and triplet excited-



state properties are strongly affected by length, structure of the solubilizing moieties, and pH of the solution. Laser flash photolysis confirmed that the NDI lowest triplet excited state was efficiently populated, upon excitation at both 355 and 532 nm, and that free amine moieties quenched both the singlet and triplet excited states by intramolecular electron transfer, with generation of detectable radical anions. Time-resolved experiments, monitoring the 1270 nm  ${}^{1}O_{2}$  phosphorescence decay generated upon laser irradiation at 532 nm, allowed a ranking of the NDIs as sensitizers, based on their  ${}^{1}O_{2}$  quantum yields ( $\Phi_{\Delta}$ ). The tetrafunctionalized **12**, exhibiting a long-lived triplet state ( $\tau \sim 32 \ \mu s$ ) and the most promising absorptivity for photodynamic therapy application, was tested as efficient photosensitizers in the photo-oxidations of 1,5-dihydroxynaphthalene and 9,10-anthracenedipropionic acid in acetonitrile and water.

# INTRODUCTION

Photodynamic therapy (PDT) is based on the use of a lightactivatable chemical, called photosensitizer (PS), and irradiation with light of appropriate wavelength to impart cytotoxicity via the generation of reactive molecular species.<sup>1–3</sup>

Typically, the useful range of wavelengths for therapeutic activation of the PS is 600–800 nm in order to optimize tissue penetration. The main process occurring upon excitation of the photosensitizer is the formation of the PS triplet excited state and energy transfer to molecular oxygen yielding singlet oxygen  $({}^{1}O_{2})$ ,<sup>4</sup> a very cytotoxic species. It is important to stress that photosensitizers that preserve their fluorescence, due to incomplete singlet–triplet intersystem crossing (ISC), can act also as fluorescent agents enabling *contemporarily* localization of the PS in the biological tissue and occurrence of the therapeutic action.<sup>5</sup> This is a very appealing objective in the framework of multimodal theranostic applications.

The triplet sensitizers actually used for PDT are limited to transition-metal complexes of porphyrinoids, reduced porphyrins, and phthalocyanines with high ISC efficiency due to the heavy atom effect of the transition metal.<sup>2,3,6–8</sup> Among their main drawbacks are low solubility and tendency to aggregate in aqueous environment leading to nonradiative decay pathways of their excited states and consequently inefficient energy transfer to oxygen. More recently, porphyrin-like photosensitizers have been included by or bound covalently to nanoparticles with size dimension in the nanometer range to optimize delivery and  ${}^{1}O_{2}$  formation.<sup>9,10</sup> Even though nanosized delivery systems for photosensitizers offer several

advantages, they suffer from drawbacks due to their complexity and often lower efficiency in the  ${}^{1}O_{2}$  generation. Therefore, the design and synthesis of water-soluble organic triplet sensitizers with intense absorption of red light, high photochemical stability, and a long-lived triplet excited state is still an interesting issue, particularly because very few have been reported yet. Recently, a bodipy-based triplet sensitizer soluble in water has been described as a singlet oxygen sensitizer exhibiting a rather low efficiency (yield in <sup>1</sup>O<sub>2</sub>, 10%).<sup>11</sup> During the last years, our attention has been focused on naphthalene diimides (NDIs) and core-substituted NDIs<sup>12</sup> as selective nucleic acid (NA) ligands and fluorescent probes. In more detail, Neidle's group and our research unit have shown that triand tetrasubstituted NDIs are potent and reversible ligands<sup>13–15</sup> as well as alkylating agents targeting guanine-rich NAs folded into G-quadruplex structures (G4s).<sup>16–19</sup> G-rich sequences able to fold into G4 are present in biologically relevant proto-oncogenes and oncogene promoters (like c-kit and  $c-myc)^{20-22}$  as well as human telomeres and participate in biological processes crucial for cell replication and survival.<sup>21,23</sup> Consequently, they represent a very appealing target and propelled the research for the development of new therapeutic approaches based on their selective targeting. Interestingly, the optoelectronic properties of the NDI core can be effectively tuned by substitution,  $^{24-26}$  as electron-rich substituents on the aromatic core give origin to strong absorption and emission in

**Received:** June 20, 2013 **Published:** July 19, 2013

the red spectroscopic window useful for PDT applications. In addition, the NDI's binding properties toward G4s<sup>14,17</sup> may also be exploited for potential selective photocleavage as suggested recently for cationic Zn-phthalocyanines.<sup>27</sup> This prompted us to start from the NDI core carrying amine substituents on the naphthalene unit to engineer efficient water-soluble  ${}^{1}O_{2}$  photosensitizers by structural modifications.

# RESULTS AND DISCUSSION

**Design and Synthesis.** In this work, we have designed and synthesized a class of water-soluble NDIs (Table 1) with unique optoelectronic properties that can be used as efficient water-soluble singlet oxygen photosensitizers and fluorescent probes.

Table 1. Structures of the Water-Soluble NDIs 1–12 Investigated as Singlet Oxygen Sensitizers

X (CH2)n	n	Y	X=NMe <sub>2</sub>	X=NMe <sub>3</sub> <sup>+</sup> Cl <sup>-</sup>
	2	Н	1	7
NH NH		Br	2	8
Y (CH <sub>2</sub> )n		$NH(CH_2)_2X$	3	9
o NO	3	Н	4	10
(CH <sub>2</sub> )n		Br	5	11
×		NH(CH <sub>2</sub> ) <sub>3</sub> X	6	12

To synthesize the above tri- and tetrasubstituted NDIs sketched in Table 1, we followed two different three-step synthetic protocols. The commercially available anhydride was brominated with dibromocyanuric acid, and the resulting product mixture containing the 2,6-dibromo-1,4,5,8-naphthale-

netetracarboxylic acid dianhydride (13) was used for a classical imidation procedure under acidic conditions yielding the 2,6-dibromo-substituted NDIs 14 and 15 (step a in Scheme 1).<sup>28,29</sup>

The following nucleophilic aromatic substitution  $(S_NAr)$  is the key step of the whole synthetic protocol. Recently, regioselectivity of the S<sub>N</sub>Ar on polybromo-substituted NDIs with aniline has been successfully achieved in the presence of fluoride.<sup>30</sup> The choice between two different conditions (b and c, Scheme 1) allows discrimination of the reactivity of the aromatic core, yielding mainly mono- (c) or difunctionalization (b). To synthetize the tetrasubstituted NDIs (3, 6) in good yields, we performed a one-pot, microwave-assisted (MW) functionalization in DMF at 135 °C for 3 min in a closed vessel. The monosubstitutions were obtained under milder conditions in acetonitrile at 70  $^{\circ}\mathrm{C}$  for 4 h using  $N^{1}\!,\!N^{1}\!$  -dimethylethane-1,2diamine or  $N^1$ ,  $N^1$ -dimethylpropane-1,3-diamine. Under these milder conditions, only regioselective monofunctionalization of the 2,6-dibromo-NDIs was efficiently achieved to give mainly the new Br derivatives 2 and 5. These Br-NDIs were prepared in order to improve intersystem crossing by the presence of the heavy atom and efficiently populate the triplet excited state.<sup>31</sup> In addition, in order to increase the lifetime of both the singlet and triplet excited states, inhibiting their deactivation by intramolecular electron transfer (eT) quenching, the corresponding quaternary ammonium salts 7-12 have been synthesized.

The quaternary ammonium salts 7-12 were synthesized by exhaustive methylation of the corresponding amines 1-6 and subsequently purified by HPLC as trifluoroacetate salts. For the NDIs embedding the ethylene spacer (1-3), the reaction carried out in acetonitrile with an excess of methyl iodide gave the desired products 7-9 with quantitative yields after 24 h at rt. Following the same strategy for the NDIs with the propylamine substituents 4-6, a mixture of partially methylated

Scheme 1. Synthesis of Water-Soluble Tri- and Tetrasubstituted NDIs<sup>a</sup>



<sup>*a*</sup>Key: (a)  $N^1$ , $N^1$ -dimethylethane-1,2-diamine (n = 2) or  $N^1$ , $N^1$ -dimethylpropane-1,3-diamine (n = 3) in acetic acid, 90 °C, 30 min, under N<sub>2</sub>; 43% yield; (b)  $N^1$ , $N^1$ -dimethylethane-1,2-diamine (n = 2) or  $N^1$ , $N^1$ -dimethylpropane-1,3-diamine (n = 3), acetonitrile, 70 °C, 4 h; (c)  $N^1$ , $N^1$ -dimethylethane-1,2-diamine (n = 2) or  $N^1$ , $N^1$ -dimethylpropane-1,3-diamine (n = 3), dry DMF, 135 °C, 3 min; MW-assisted and closed vessel synthesis; (d) MeI, acetonitrile, rt, 16 h.

quaternary ammonium salts was obtained. In order to get the NDIs 10-12 a suspension of sodium carbonate was required to achieve quantitative conversion. Such a difference in reactivity is likely the result of a different basicity of the amines. All the NDIs 1-12 have been purified by HPLC using ACN/H<sub>2</sub>O + 0.1% CF<sub>3</sub>COOH and characterized as chloride salts.

**Photophysical Characterization.** Parts a and b of Figure 1 show the absorption and corrected fluorescence spectra of the NDI quaternary ammonium salts 7-12 (Scheme 1).



Figure 1. UV-vis absorption (solid lines) and fluorescence spectra (dotted lines) of the NDIs (a) 7–9 and (b) 10–12. Excitation at 485 nm for 7, 8, 10, and 11 and 600 nm for 9 and 12 (1 cm path length).

The absorption spectra of the amines 1-6 in solutions of different pH are very similar to those of the quaternary ammonium salts, so the protonation state of the terminal amine groups (NMe<sub>2</sub>) does not influence the absorption spectra (see the Supporting Information). The absorption band with vibronic signature in the 300–400 nm range is typical of the NDI core and is not substantially affected by substitution.<sup>32</sup> It clearly emerges how the introduction of just one amine substituent is able to generate a second absorption band in the visible region of comparable intensity to that with a maximum at 370 nm.

Compared to the H-derivative, bromination of the NDI core shifts the absorption band in the visible spectrum to the red by ca. 10 nm due to the Br atom acting as weak electron-donating substituent. The most significant shift of ca. 90 nm is obtained by introduction of the second amine substituent on the NDI core. With the introduction of the second amine substituent the NDI derivative complies with the PDT requirements of strong absorption in the 600–800 nm spectroscopic window. As underlined in several studies, these interesting electronic properties arise from a charge transfer (CT) transition involving the doublet of the aromatic amines.<sup>25,33</sup>

Table 2 collects the photophysical data of all the quaternary ammonium salts 7-12, in water, and their related amines (1-6), under acid buffered conditions. Measurements were carried out under acidic conditions in an attempt to protonate all

Table 2. Photophysical Properties of the NDI Compounds in Water for the Salt Derivatives in Phosphate Buffer of pH 2 for the Neutral Derivatives

NDIs	$\lambda_{\max} (nm)$	$\varepsilon_{\rm max}({ m M}^{-1}~{ m cm}^{-1})$	$\Phi_{\mathrm{F}}{}^{a}$	$ au_{\mathrm{f}}  (\mathrm{ns})^b$	$\Phi_{\Delta}{}^c$	$ au_{\mathrm{T}} \ (\mu \mathrm{s})^d$
4	522	11000	0.19	5.60		
10	522	11000	0.21	5.50		0.34
1	509	7100	0.29	7.36		
7	511	7100	0.29	7.35		0.18
6	616	7400	0.17	4.40		
12	613	10700	0.17	4.00	0.30	32
3	598	13000	0.26	7.00	0.04	
9	598	13000	0.27	7.10	0.07	7.4
5	531	6400	0.11	3.50	0.39	26
11	530	7600	0.12	3.33	0.46	22
2	518	7200	0.11	3.40	0.48	30
8	517	7300	0.11	3.00	0.63	23

<sup>*a*</sup>Using Ru(bpy)<sup>3+</sup> as reference with a value of 0.028 in aerated water for Br and H derivatives and compound 11 in water as reference for 3, 6, 9, and 12. <sup>*b*</sup>Excitation at 373 nm. <sup>*c*</sup>Determined from phosphorescence of <sup>1</sup>O<sub>2</sub> at 1270 nm in D<sub>2</sub>O, under air-equilibrated conditions. All solutions are isosbestic at the 532 nm excitation wavelength. <sup>*d*</sup>Triplet lifetime in argon-saturated solution. Excitation at 532 nm.

terminal amine groups. Protonation of the aromatic amines was negligible at pH above 1, as the absorption spectra did not change (see the Supporting Information).

As expected, the lowest fluorescence quantum yields and the shortest lifetimes have been observed for the Br derivatives. In fact, the presence of the bromide substituent directly on the NDI core lowers both parameters due to the heavy atom effect. Concerning the tri- and tetraderivatives, we observe an unexpected effect of the length of the alkyl chain. In fact, the ethyl derivatives (1, 3, 7, and 9) have better fluorescence performance compared to the propyl derivatives (4, 6, 10, and 12). Considering the quaternary ammonium salts having all aliphatic amines positively charged and not available for eT processes, one could conclude that the nonradiative deactivation processes are more efficient for compounds with the longer and more flexible alkyl spacer (n = 3), as the fluorescence lifetimes are significantly reduced compared to those of the compounds with shorter spacer (n = 2, see Table)2). Indeed, radiative rate constants  $(k_r = \Phi/\tau)$  are very similar for H and tetraderivatives, while the rate constant for nonradiative processes of the compounds with a long spacer is twice that of the compounds with a shorter spacer. The fluorescence spectra of 5-7 at three different pH values are reported in Figures 4S-6S (Supporting Information). At pH 7 we observe lower relative  $\Phi_F$  values, suggesting deprotonation of an ammonium moiety and subsequent quenching of the emitting singlet excited state by intramolecular eT. The fluorescence decay is monoexponential at all pH values, and the lifetimes values do not vary significantly. For a quantitative evaluation of the acidity of the ammonium moieties and the remarkable effect of the spacer length on it, fluorescence and potentiometric titrations have been performed.

**Evaluation of the Ground-State NDIs' Acidity by Fluorescence and Potentiometric Titrations.** The mode of protonation of the solubilizing amino moieties tethered to the NDI core was investigated by potentiometric and fluorescence titrations, as it controls the quenching of the excited states by eT. The NDIs 3 and 6 were chosen as models, as their fluorescence quantum yields dropped passing from pH 2 to pH 7. Fluorescence intensity change as a function of pH was analyzed for both amines (Figure 2).



Figure 2. Fluorescence intensity ( $I_f$ ) vs pH. The sigmoids defined by the (a) empty squares and (b) black circles describe the fluorescent quenching of 3 (at 640 nm) and 6 (at 661 nm), respectively, both 5 × 10<sup>-6</sup> M in aqueous solution, 0.1 M in NaNO<sub>3</sub>.

The resulting sigmoids clearly suggest that the deprotonation occurs at a lower pH for 3. The absorption spectra of the NDIs 3 and 6 in water solutions are very similar at different pH values, so the protonation state of the terminal amine groups does not influence the absorption spectra at  $pH \leq 9$ . Under more basic conditions (pH  $\geq$  9), the amine 3 exhibited an additional absorption band at longer wavelength ( $\lambda_{max}$  625 nm) than that observed under acid and neutral conditions ( $\lambda_{max}$  597 nm). These spectroscopic data suggest an aggregation of 3 in dimer and/or oligomer aggregates, rather than a deprotonation of the aromatic amine, as the latter has never been reported on similar NDIs in water. As at pH > 10 precipitation of 3 occurred, we were unable to measure its  $pK_{a4}$ . Therefore, the potentiometric titration of 3 yielded only three  $pK_a$  values:  $pK_{a1}$ = 5.6,  $pK_{a2}$  = 7.4, and  $pK_{a3}$  = 7.7, suggesting that when the amine moiety is fairly close to the NDI core, due to a short alkyl spacer, the ammonium salts are rather acidic. In fact, the NDI 6 was much more basic and did not show any detectable aggregation into dimers or oligomers at pH  $\leq$  12. Therefore, we were able to measure all the  $pK_a$  constants for 6 ( $pK_{a1} = 8.1$ ,  $pK_{a2} = 8.6$ ,  $pK_{a3} = 8.8$ , and  $pK_{a3} = 11.3$ ).

The speciation analysis in Figure 3, resulting from the potentiometric titrations, describes the distribution of tetracationic vs tri-, bis-, and monocationic and neutral NDIs (NDI-H<sub>4</sub>, NDI-H<sub>3</sub>, NDI-H<sub>2</sub>, NDI-H, and NDI, respectively) as function of the pH. These data suggest that the amine **6** is mainly fully protonated at pH 7 (94% of **6**-H<sub>4</sub>), unlike the amine **3**, which exists as a mixture of both tri- (68%, **3**-H<sub>3</sub>) and dicationic (26%, **3**-H<sub>2</sub>) species.

These data clearly indicate the NDI **6** is a better fluorescent probe than **3** under physiological conditions; being fully protonated, it is not quenched by intramolecular eT. Surprisingly, the fluorescence quenching of **3** occurs during the deprotonation of the second ammonium moiety in the species **3**-H<sub>3</sub>, probably because the first free amino group is not enough electron donor due to the closer NDI core. On the contrary, the fluorescence quenching of **6** follows the deprotonation of the tetracationic species **6**-H<sub>4</sub>.



**Figure 3.** Speciation analysis resulting from the potentiometric titrations for the NDIs (a) **3** (tetracationic **3**- $H_4$ , tricationic **3**- $H_3$ , and dicationic **3**- $H_2$  distribution is reported) and (b) **6** (the distribution of **6**- $H_4$ , **6**- $H_3$ , **6**- $H_2$ , **6**- $H_3$ , and **6** are reported). The diamonds describe the fluorescent quenching.

Spectroscopic and Kinetic Characterization of the NDI Triplet State. To provide a full characterization of the synthesized NDI as singlet oxygen sensitizers, it is mandatory to examine the key features of their triplet states. In fact, both the population of the triplet excited states and their lifetimes have a direct effect on the efficiency for singlet oxygen production. Therefore, we set out to examine these issues in detail for several of our NDIs in acetonitrile and in water solution under neutral and slightly acidic conditions (pH 5 and 7), by nanosecond laser flash photolysis using a Nd:YAG laser, operating at both 532 and 355 nm. Upon laser excitation at both 355 and 532 nm, transient absorption spectra in the range of 380–750 nm, with  $\lambda_{\rm max}$  at 430 nm, were observed for all the substituted NDIs investigated. A second broad band exhibiting  $\lambda_{\rm max}$  centered at 530, 630, and 670–690 nm was recorded for the monosubstituted (1 and 4), Br-substituted (2 and 5), and tetrasubstituted NDIs (9 and 12) (Figures 4 and 5).



Figure 4. Time-resolved difference absorption spectra flashing a  $5 \times 10^{-5}$  M solution of 1, 2 (at 355 nm), 3 (at pH 5), and 9 (at pH 7), both at 532 nm, in argon-purged  $10^{-2}$  M phosphate buffer.



**Figure 5.** Time-resolved difference absorption spectra flashing a 5  $\times$  10<sup>-5</sup> M solution of 4, 5 (at 355 nm), 6 (at pH 5), and 12 (at pH 7), both at 532 nm, in argon-purged 10<sup>-2</sup> M phosphate buffer.

It is likely that the electron-donor character of the second substituent on the NDI core, passing from H, Br to NHR, resulted in a progressive red shift of the second band, which also became more intense. Bleaching at 510-550 nm and 550-600 nm for the NDIs 1, 2, 4, 5 and 3, 6, 9, 12, respectively, was due to the depletion of the ground state of the NDIs upon excitation to the triplet excited state. All transient absorption profiles monitored at both 430 and 680 nm decay monoexponentially in air-equilibrated solutions with a lifetime of ~2  $\mu$ s (Figure 6).

In argon-saturated solutions, lifetimes increase to ca. 20-30  $\mu$ s, indicating that at pulse end we are very likely observing the triplet-state absorption. The transient absorption spectra in Arsaturated solution observed from the pulse end to 80  $\mu$ s after (Figures 3 and 4) have been assigned to the triplet state of each NDI, on the basis of its sensitivity to O<sub>2</sub> and similarity to the  $T_1 - T_n$  absorption of non-water-soluble NDIs.<sup>34</sup> The amines 3 and 6 exhibit a fairly similar spectra at pulse end to their quaternary ammonium analogues 9 and 12. Nevertheless, at pH 5, both the amines 3 and 6 reveal the generation of a more stable species on a longer time scale (>200  $\mu$ s). This second species, which was completely bleached by oxygen, also exhibits an absorption spectrum ( $\lambda_{max}$  480–490 and 660 nm) fairly similar to the stable species generated by NDI monoelectronic reduction. Therefore, it can be assigned to the NDI radical anions  $3^{\bullet-}$  and  $6^{\bullet-}$  generated by eT from free amine to the excited NDI core.<sup>35–37</sup> In the hypothesis that the NDIs 3 and 9 and the homologues 6 and 12 have similar molar absorption coefficients for T-T absorption at 430 nm, we see that initial absorbance is much higher for 12 compared to the other tetrasubstituted compounds 3, 6, and 9 (Figure 6). This allows us to conclude that the triplet is likely formed with much higher efficiency in the case of 12. It is somewhat surprising that its analogue 6 at pH 2 did not efficiently produce the triplet excited state. In spite of very similar fluorescence properties of 6 and 12, the triplet formation yield seems much lower. A possible explanation for this behavior may be the presence of the carbonyl group enabling excited state intramolecular proton transfer (ESIPT) from the protonated aliphatic amine to the carbonyl.<sup>38</sup>

**Singlet Oxygen Generation Efficiency.** The fluorescence data are in agreement with the ability of these compounds to



**Figure 6.** Differential absorption decay monitored at (a) 430 nm and (b) 660 nm for **3**, **6**, **9**, and **12**  $(8 \times 10^{-5} \text{ M})$  and (c) 430 nm for **2**, **5**, **8**, and **11**  $(4 \times 10^{-5} \text{ M})$ , in air-equilibrated neutral D<sub>2</sub>O solution (9, **12**, **8**, and **11**) or acidic D<sub>2</sub>O of pH 2 (**3**, **6**, **2**, and **5**). All of the solution were excited at 532 nm where they exhibited identical absorbances of 0.3.

produce singlet oxygen. The singlet oxygen quantum yields  $(\Phi_{\Delta})$  reported in the present study, and listed in Table 2, were obtained in time-resolved experiments by comparing the initial intensity of the 1270 nm singlet oxygen phosphorescence decay (Figure 7) produced upon irradiation at 532 nm (by means of a Nd:YAG laser) of the given NDIs to that of 5,10,15,20-tetrakis(4-sulfonatophenyl)porphyrin (TPPS) as standard sensitizer ( $\Phi_{\Delta} = 0.76$ ).<sup>39,40</sup>

As expected, the Br derivatives (2, 5, 8, and 11) produce  ${}^{1}O_{2}$  in water. Our approach to introduce a Br substituent favoring intersystem crossing thus gave positive results. The  $\Phi_{\Delta}$  values are good ranging from 0.39 to 0.63, even though they are lower compared to the TPPS value. From the point of view of application in PDT, their absorption extending up to 550 nm represents, however, a drawback. Under the very same experimental conditions, the Br derivatives exhibit higher  $\Phi_{\Delta}$  than that of the another conventional photosensitizer Rose Bengal (Figure 7). Such a difference may be due to aggregation and T–T annihilation known to occur for Rose Bengal.

More surprisingly, the quaternary ammonium salt 12 was the only NDI among those without Br, able to produce singlet oxygen in water with a very satisfactory yield (0.30). Such an



Figure 7. Time-resolved phosphorescence signal of singlet oxygen in  $D_2O$  (monitored at 1270 nm), using the standards TPPS and Rose Bengal and the NDIs 2, 3, 5, 8, 9, 11, 12 as photosensitizers. For the amines 2, 3, and 5 the pH was set to 2 by DCl addition. Excitation with Nd:YAG laser at 532 nm; energy of 3.6 mJ/pulse; all solutions absorb equally at 532 nm.

efficiency was even more striking in comparison to the very low  $\Phi_{\Delta}$  (0.07) for its analogue 9, exhibiting a shorter alkyl spacer (n = 2). The related ethyl- (n = 2) and propylamino (n = 3) analogues 3 and 6 produce singlet oxygen with negligible yields ( $\leq 0.04$ ), even at pH 2, which ensures complete protonation of all the amino moieties and erases the excited state quenching by intramolecular eT. As suggested above, this behavior may be due to the presence of the carbonyl group enabling excited-state intramolecular proton transfer from the protonated aliphatic amine to the carbonyl.<sup>38</sup> This, however, does not explain the poor behavior of compound 9 with respect to that of 12.

Photooxidation of 1,5-Dihydroxynaphthalene and 9,10-Anthracenedipropionic Acid in Acetonitrile and Water by the Tetracationic NDI 12. Sensitizers such as 12, which are characterized by (i) intense absorption of red visible light, (ii) long-living triplet, and (iii) high singlet oxygen yields are ideal for performing efficient photo-oxidations. In addition, the solubility of 12 in both acetonitrile and water allows to exploit its properties as singlet oxygen sensitizer in both solvents, also targeting water-soluble biomolecules. Thus, as a proof of concept, we decided to investigate 12 as singlet oxygen sensitizer in the photo-oxidation of 1,5-dihydroxynaphthalene (DHN) and 9,10-anthracenedipropionic acid (ADPA) in acetonitrile and water, respectively. Visible light at longer wavelength than 500 nm was used. DHN was efficiently oxidized to its naphthoquinone derivative (5-hydroxy-1,4naphthalenedione) through the endoperoxide, by <sup>1</sup>O<sub>2</sub>.<sup>41</sup> The resulting naphthoquinone absorbing at 360-440 nm, has been efficiently monitored by the increase of its absorption, which appeared to be in the spectroscopic window where the NDI shows negligible absorption (Figure 8a).

Unfortunately, the solubility in water of DHN is very poor, and we were able to perform a photo-oxidation of DHN only in aqueous acetonitrile (Supporting Information). Therefore, DHN has been replaced by ADPA as water-soluble  ${}^{1}O_{2}$  trap.<sup>42</sup> Contrary to DHN, in this case, the photo-oxidation has been followed by the decrease absorption at 400 nm due to the consumption of ADPA anthracene chromophore, forming a colorless endoperoxide (Figure 8b). Continuous photobleaching of the anthracene absorption was observed for 10 h by using excitation wavelength longer than 450 nm. Byproduct formation was not detected by UV absorption during the



**Figure 8.** Change in the absorption spectra by irradiation (at  $\lambda > 450$  nm) of (a) an acetonitrile solution of DHN ( $1.2 \times 10^{-4}$  M) and (b) an aqueous solution of ADPA ( $1.2 \times 10^{-4}$  M), using **12** ( $2 \times 10^{-5}$  M and  $6 \times 10^{-5}$  M, respectively) as singlet oxygen photosensitizer.

irradiation. No decrease in the anthracene absorbance was observed in the solutions without the sensitizer. To our knowledge, this is the first fairly efficient photobleaching experiments of ADPA, performed in H<sub>2</sub>O, rather than in D<sub>2</sub>O, where the lifetime of singlet oxygen is much longer. This evidence further suggests that **12** is a promising singlet oxygen sensitizer for biological applications. A small blue shift (6 nm) of the absorption at 640 nm was detected during the photooxidation, suggesting a weak interaction between ADPA and the NDI core, in water solution.

## CONCLUSIONS

Naphthalene diimide derivatives with bromo and amino or quaternary ammonium substituents on both the aromatic core and the imide moieties were prepared as water-soluble singlet oxygen sensitizers, in which water solubility derived from ionic substituents. Their photophysical properties were thoroughly investigated by steady-state and time-resolved spectroscopy. The red colored Br-containing NDIs (2, 5, 8, and 11) were very efficient singlet oxygen sensitizers in water solution. Unfortunately, their absorbing wavelength was centered at 537 nm, slightly below the PDT spectroscopic window. Differently, the blue colored tetrasubstituted NDI 12 retains a comparable efficiency in the generation of  ${}^{1}O_{2}$ , coupled to an absorption in the red. Moreover, all of the NDI investigated, including the best singlet oxygen sensitizers, also exhibited a remarkable red (for the Br-NDIs 2, 5, 8, and 11) and NIR (tetra-substituted NDIs 3, 6, 9, and 12) fluorescence emission with lifetimes that are longer than those of intrinsic biological fluorophores. This

Article

evidence makes them very appealing candidates for multimodal applications as fluorescent reporters and efficient photosensitizers for PDT. Moreover, taking into consideration that cationic NDIs have already been proved to be excellent and selective G-quadruplex ligands, NDIs such as **12** may be exploited as selective G4 ligands as well as agents to achieve photocleavage footprinting of G-quadruplex folding nucleic acids. These aspects are currently under investigation.

#### EXPERIMENTAL SECTION

Synthesis and Purification. The NDIs 1, 3, 4, and 6 have been already synthesized and characterized.<sup>14</sup> In this study, they have been synthesized following a more efficient protocol, starting from the precursors 14 and 15. The anhydride 13 and the NDIs 14 and 15 have been synthesized according to published procedures.<sup>17,19</sup> HPLC analysis and purifications were performed using both preparative and analytical HPLCs. The analytical column was XSelect CSH Phenyl-Hexyl (150  $\times$  4.6 mm). The preparative column was XSelect CSH Prep Phenyl-Hexyl 5  $\mu$ m (150 × 30 mm). Flows were 1 mL/min for analytical and 27 mL/min for preparative. An analytical method was used: method A (Aqueous solvent: 0.1% trifluoroacetic acid in water; organic solvent: acetonitrile; gradient: 95% aqueous, gradually to 0% aqueous over 14 min and at the end an isocratic flow over 2 min). Preparative HPLC was performed using method B (aqueous solvent: 0.1% trifluoroacetic acid in water; organic solvent: acetonitrile; gradient: 95% aqueous, gradually to 20% aqueous over 18 min and at the end an isocratic flow over 4 min). <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on a 300 MHz spectrometer, and the chemical shifts are reported relative to TMS. The structures of new compounds were deduced from the results of <sup>1</sup>H and <sup>13</sup>C NMR.

Nucleophilic Aromatic Substitution Reaction at rt, Optimized for Br-NDI Synthesis. The NDI 14–15 (0.5 mmol) was dissolved into 40 mL of acetonitrile in a round-bottom flask together with the corresponding amine (*N*,*N*-dimethylethylamine for 1 and 2 and *N*,*N*-dimethylpropylamine for 4 and 5; 1.5 mmol). The mixture was stirred at 70 °C for 4 h under argon. The resulting red solution was concentrated under vacuum and a red solid was obtained. The crude product was purified by preparative HPLC chromatography, using a C-18 reversed-phase column (CH<sub>3</sub>CN/H<sub>2</sub>O 0.1%TFA) according to analytical method B. HCl 1 M solution was added to each chromatographic portion. Solvent evaporation under vacuum at rt afforded the adducts 1 (99.4 mg, 33% yield), 2 (142.8 mg, 42%), 4 (93.7 mg, 29%), and 5 (163.1 mg, 45%) as hydrochlorides.

*N*,*N*<sup>'</sup>-Bis((dimethylamino)ethylamino)-2-bromo-6-((dimethylamino)ethylamino)-1,4,5,8-naphthalenetetracarboxylic Bisimide Trihydrochloride (2·3HCl). The collected solid was purified by preparative HPLC chromatography (CH<sub>3</sub>CN/H<sub>2</sub>O 0.1%TFA). Red solid. Mp > 350 °C dec. <sup>1</sup>H NMR (300 MHz, D<sub>2</sub>O): 8.68 (s, 1H), 8.3 (s, 1H), 4.74 (m, 4H), 4.38 (m, 2H), 3.96 (m, 2H), 3.75 (m, 4H), 3.41 (bs, 18H). <sup>13</sup>C NMR (75 MHz, D<sub>2</sub>O): 165.1; 162.3; 161.6; 151.1; 137.6; 127.9; 126.7; 122.7; 121.1; 120.8, 120.4; 100.7; 64.1; 62.0; 61.7; 53.6; 53.3; 36.9; 34.7; 34.0. Anal. Calcd for C<sub>26</sub>H<sub>36</sub>BrCl<sub>3</sub>N<sub>6</sub>O<sub>4</sub>: C, 45.73; H, 5.31; N, 12.31. Found: C, 45.82; H, 5.29; N, 12.24.

*N*,*N*′-Bis((dimethylamino)propylamino)-2-bromo-6-((dimethylamino) propylamino)-1,4,5,8-naphthalenetetracarboxylic Bisimide Trihydrochloride (5·3HCl). The collected solid was purified by preparative HPLC chromatography (CH<sub>3</sub>CN/H<sub>2</sub>O 0.1% TFA). Red solid. Mp > 350 °C dec. <sup>1</sup>H NMR (300 MHz, CD<sub>3</sub>OD): 8.21 (s, 1H); 8.00 (s, 1H); 4.15 (m, 4H); 3.76 (t, <sup>3</sup>*J* = 7.6 Hz, 2H); 3.43 (m, 2H); 3.31 (m. 4H); 3.27 (s, 6H); 2.94 (s, 12H); 2.34 (m, 2H); 2.17 (m, 4H). <sup>13</sup>C NMR (75 MHz, CD<sub>3</sub>OD): 166.8; 163.1; 162.4; 152.8; 138.4; 129.3; 128.3; 124.1; 123.9; 122.3; 121.5; 121.2, 101.0; 57.0; 56.9; 56.7; 43.8; 41.4; 39.5; 38.8; 26.1; 24.8; 24.7. Anal. Calcd for C<sub>29</sub>H<sub>42</sub>BrCl<sub>3</sub>N<sub>6</sub>O<sub>4</sub>: C, 48.05; H, 5.84; N, 11.59. Found: C, 47.95; H, 5.82; N, 11.66.

General Procedure for the Microwave-Assisted (MW) Nucleophilic Aromatic Substitution, Optimized for NDIs Containing no Br. The NDI 14–15 (0.5 mmol) was dissolved into 6 mL of dry DMF in a MW vial together with the corresponding amine (*N*,*N*-dimethylethylamine for **3** and *N*,*N*-dimethylpropylamine for **6**; 1.5 mmol). The mixture was stirred and heated in a microwave reactor, according a closed vessel protocol, at 135 °C, 200 psi, 200 W, for 3 min. The resulting dark-violet solution was cooled at rt, and water (50 mL) was added to induce precipitation. The filtered crude product was purified by preparative HPLC chromatography, using a C-18 reversed-phase column (CH<sub>3</sub>CN/H<sub>2</sub>O 0.1%TFA, method B). HCl 1 M solution was added to each chromatographic portion. Solvent evaporation under vacuum afforded the adducts **1** (152.5 mg, 51%, yield), **3** (97.8, 27%), **4** (148.6 mg, 46%), and **6** (125.2 mg, 32%) as hydrochlorides.

**Exhaustive Methylation of the NDI 1–5.** The NDIs purified as hydrochlorides were dissolved in a NaHCO<sub>3</sub> solution and extracted three times with  $CH_2Cl_2$ . The recovered organic layers were dried on Na<sub>2</sub>SO<sub>4</sub>, and the solvent was evaporated under reduced pressure. The collected amine (0.5 mmol) was suspended in 50 mL of  $CH_3CN$ , and 0.24 g (1.7 mmol) of  $CH_3I$  was added. This suspension was stirred under nitrogen atmosphere for 12 h. After this period, the solvent was removed under vacuum, and the crude was purified by preparative HPLC, using a C-18 reversed-phase column ( $CH_3CN/H_2O$  0.1%TFA, method B). HCl 1 M solution was added to each chromatographic portion. Solvent evaporation under vacuum at rt afforded the adducts to give the quaternary ammonium salts as chlorides 7–11.

*N*,*N*′-Bis((trimethylamino)ethylamino)-2-((trimethylamino)ethylamino)-1,4,5,8-naphthalenetetracarboxylic Bisimide Trichloride (7). Red solid, 309 mg. Yield: 96%. Mp > 350 °C. <sup>1</sup>H NMR (300 MHz, D<sub>2</sub>O): 8.59 (d, <sup>3</sup>*J* = 7.9 Hz, 1H); 8.33 (d, <sup>3</sup>*J* = 7.9 Hz, 1H); 8.24 (s, 1H); 4.63 (m, 4H); 4.27 (t, <sup>3</sup>*J* = 6.5 Hz, 2H); 3.84 (t, <sup>3</sup>*J* = 6.5 Hz, 2H); 3.71 (m, 4H); 3.31 (s, 27H). <sup>13</sup>C NMR (75 MHz, D<sub>2</sub>O): 165.7; 164.3; 164.1; 163.7; 151.8; 131.5; 129.2; 127.9; 125.9; 125.2; 123.2; 119.8; 118.7, 101.0; 64.2; 62.3; 62.0; 53.7; 53.3; 36.8; 34.3; 33.9. Anal. Calcd for C<sub>29</sub>H<sub>43</sub>Cl<sub>3</sub>N<sub>6</sub>O<sub>4</sub>: C, 53.91; H, 6.71; N, 13.01. Found: C, 54.03; H, 6.70; N, 13.11.

*N*,*N*'-Bis((trimethylamino)ethylamino)-2-bromo-6-((trimethylamino)ethylamino)-1,4,5,8-naphthalenetetracarboxylic Bisimide Trichloride (8). Red solid, 325 mg. Yield: 90%. Mp > 350 °C. <sup>1</sup>H NMR (300 MHz, D<sub>2</sub>O): 8.53 (s, 1H); 8.19 (s, 1H); 4.59 (m, 4H); 4.25 (t, <sup>3</sup>*J* = 6.3 Hz, 2H); 3.84 (t, <sup>3</sup>*J* = 6.3 Hz, 2H); 3.62 (m, 4H); 3.29 (s, 27H). <sup>13</sup>C NMR (75 MHz, D<sub>2</sub>O): 165.0; 162.2; 161.6; 151.0; 137.5; 127.9; 126.7; 122.6; 121.1; 120.8; 120.3; 100.6; 63.9; 61.8; 61.6; 53.5; 53.2; 36.8; 34.6; 33.9. Anal. Calcd for  $C_{29}H_{42}BrCl_3N_6O_4$ : C, 48.05; H, 5.84; N, 11.59. Found: C, 48.22; H, 5.78; N, 11.35.

N, N' - B is ((trimethylamino)-1,4,5,8-naphthalenetetracarboxylic Bisimide Tetrachloride (9). Violet solid, 386 mg. Yield: 99%. Mp > 350 °C. <sup>1</sup>H NMR (300 MHz, D<sub>2</sub>O): 8.10 (s, 2H); 4.59 (m, 4H); 4.11 (m, 4H); 3.71 (m, 4H); 3.59 (m, 4H); 3.21 (s, 36H). <sup>13</sup>C NMR (75 MHz, D<sub>2</sub>O): 167.4; 165.4; 164.6; 164.1; 150.0; 127.2; 123.1; 119.7; 115.9; 104.6; 65.7; 63.7; 55.0; 54.8; 38.0; 35.4. Anal. Calcd for C<sub>34</sub>H<sub>56</sub>Cl<sub>4</sub>N<sub>8</sub>O<sub>4</sub>: C, 52.18; H, 7.21; N, 14.32. Found: 52.31; H, 7.18; N, 14.27.

*N*, *N*′ - Bis ( (trimethylamino)-1,4,5,8-naphthalenetetracarboxylic Bisimide Trichloride (10). Red solid, 337 mg. Yield: 98%. Mp > 350 °C. <sup>1</sup>H NMR (300 MHz, D<sub>2</sub>O): 8.2 (d, <sup>3</sup>J = 7.9 Hz, 1H); 7.97 (d, <sup>3</sup>J = 7.9 Hz, 1H); 7.79 (s, 1H); 4.11 (m, 4H); 3.68 (t, 2H); 3.46 (m, 6H), 3.10 (s, 9H); 3.06 (s, 18H); 2.16 (m, 6H). <sup>13</sup>C NMR (75 MHz, D<sub>2</sub>O):  $\delta$ =165.3; 163.8; 163.6; 163.3; 151.7; 130.8; 128.6; 127.0; 125.1; 124.4; 122.3; 119.6; 118.6, 99.5; 63.9; 52.9; 39.5; 37.7; 37.1; 22.9; 21.4. Anal. Calcd for C<sub>32</sub>H<sub>49</sub>Cl<sub>3</sub>N<sub>6</sub>O<sub>4</sub>: C, 55.85; H, 7.18; N, 12.21. Found: C, 55.71; H, 7.27; N, 12.30.

*N*,*N*'-Bis((trimethylamino)propylamino)-2-bromo-6-((trimethylamino)propylamino)-1,4,5,8-naphthalenetetracarboxylic Bisimide Trichloride (11). Red solid, 372 mg. Yield: 97%. Mp > 350 °C. <sup>1</sup>H NMR (300 MHz, D<sub>2</sub>O): 8.18 (s, 1H); 7.92 (s, 1H); 4.05 (m, 4H); 3.68 (m, 2H); 3.49 (m, 2H); 3.38 (m, 4H), 3.10 (s, 9H); 3.03 (s, 18H); 2.24 (m, 2H); 2.10 (m, 4H). <sup>13</sup>C NMR (75 MHz, D<sub>2</sub>O): 165.0; 162.2; 162.0; 161.3; 151.2; 137.0; 127.5; 126.1; 121.9; 120.4; 119.9; 118.0; 114.1, 99.2; 63.8; 52.8; 39.6; 38.1; 37.2; 22.7;

21.2; 21.1. Anal. Calcd for  $C_{32}H_{48}BrCl_3N_6O_4$ : C, 50.11; H, 6.31; N, 10.96. Found: C, 49.89; H, 6.35; N, 11.03.

General Protocol for Exhaustive Methylation of Tetrasubstituted NDI 6. The NDI 6 purified as hydrochloride was dissolved in a Na<sub>2</sub>CO<sub>3</sub> solution and extracted three times with  $CH_2Cl_2$ . The recovered organic layers were dried on Na<sub>2</sub>SO<sub>4</sub>, and the solvent was evaporated under reduced pressure. The collected amine (0.5 mmol) was added to a suspension of Na<sub>2</sub>CO<sub>3</sub> (2 mmol) in 50 mL of CH<sub>3</sub>CN, and 0.24 g (1.7 mmol) of CH<sub>3</sub>I was added. This suspension was stirred under nitrogen atmosphere for 12 h. After this period, the solvent was removed under vacuum, and the crude was purified by preparative HPLC using a C-18 reversed-phase column (CH<sub>3</sub>CN/H<sub>2</sub>O 0.1%TFA, method B). HCl 1 M solution was added to each chromatographic portion. Solvent evaporation under vacuum at rt afforded the adducts to give the quaternary ammonium salt as chloride **12** (415 mg, 99%, yield).

N, N' - B is ((trimethylamino) propylamino)-1,4,5,8-naphthalenetetracarboxylic Bisimide Tetrachloride (12). Violet-dark solid. Yield: 99%. Mp > 350 °C. <sup>1</sup>H NMR (300 MHz, D<sub>2</sub>O): 7.62 (s, 2H); 4.14 (m, 4H); 3.62 (m, 4H); 3.49 (m, 8H); 3.08 (s, 18H); 3.06 (s, 18H); 2.17 (m, 8H). <sup>13</sup>C NMR (75 MHz, D<sub>2</sub>O): 165.3; 163.3; 148.2; 124.6; 120.5; 117.3, 101.6; 64.1; 53.1; 39.2; 37.3; 22.7; 21.6. Anal. Calcd for C<sub>38</sub>H<sub>64</sub>Cl<sub>4</sub>N<sub>8</sub>O<sub>4</sub>: C, 54.41; H, 7.69; N, 13.36. Found: C, 54.22; H, 7.79; N, 13.26.

Sample Preparation for Solution Studies. For the spectroscopic measurements we used pure water as well as  $10 \text{ mM K}^+$  phosphate buffer of pH 2 adjusted with aliquots of HCl 3 N.

Absorption and Fluorescence Spectra. UV-vis absorption spectra were recorded on a standard commercial spectrophotometer. Fluorescence spectra were measured using 1 nm steps and 0.5 s dwell time. Slits were kept narrow to 1 nm in excitation and 1 or 2 nm in emission. Where necessary, a cutoff filter was used. Right angle detection was used. All the measurements were carried out at 295 K in quartz cuvettes with path length of 1 cm. All fluorescence spectra have been obtained for air-equilibrated solutions absorbing less than 0.1 at all wavelengths to avoid inner filter effects and reabsorption of emission. Furthermore, they have been corrected for wavelength dependent response of the monochromator/PMT couple. The compound Ru(bpy)<sub>3</sub>Cl<sub>3</sub>.xH<sub>2</sub>O dissolved in air-equilibrated water with known fluorescence quantum yield ( $\Phi_{\rm F}$ ) of 0.028 was used as standard for the determination of the fluorescence quantum yield of the NDI samples. The  $\Phi_{\rm F}$  value obtained for the compound 11 in water was used as reference to determine the fluorescence quantum yield of the tetra-NDI compounds (3, 6, 9 and 12), excited at 546 nm. Using the same solvents for all compounds and iso-absorbing solutions at the excitation wavelength no corrections had to be made for absorbance neither solvent refraction index and we calculated the fluorescence quantum yields,  $\Phi_{\rm F}$ , using the formula below, with A being the integrated area of the corrected fluorescence spectra:

$$\Phi_{\rm F} = \Phi_{\rm F}^{\rm ref} \times A/A^{\rm ref} \tag{1}$$

**Fluorescence Lifetimes.** They were measured in air-equilibrated solutions with a time correlated single photon counting system. A nanosecond LED source at 373 nm was used for excitation and the emission was collected at right angle at 580 or 635 nm using a long pass cutoff filter at 495 nm. Decay profiles were fitted using a mono- or multiexponential function and deconvolution of the instrumental response.

$$I(t) = \Sigma_{i}a_{i} \times \exp(-t/\tau_{i})$$
<sup>(2)</sup>

$$f_{i} = (a_{i} \times t_{i}) / \Sigma_{j} (a_{j} \times t_{j})$$
<sup>(3)</sup>

**Nanosecond Laser Flash Photolysis.** The pulse of a Nd:YAG laser, operating at 355 and 532 nm (20 ns fwhm, 2 Hz), was suitably shaped passing through a 3 mm high and 10 mm wide rectangular window and providing a fairly uniform energy density of 2.7 mJ/pulse corresponding to 9 mJ/cm<sup>2</sup> incident on the sample cell. A front portion of 2 mm of the excited solution was probed at right angle, the useful optical path for analyzing light being 10 mm.  $A_{532}$  was ~0.3 over

1 cm. Ar-saturated or air-equilibrated solutions were used. The sample was renewed after few laser shots. Temperature was 295 K.

Singlet Oxygen Time-Resolved Emission Measurements. The pulse of a Nd:YAG laser, operating at 532 nm (20 ns fwhm, 2 Hz), was used for excitation of the reference TPPS and the NDI samples dissolved in air-equilibrated deuterated water. We used DCl to adjust pH values to 2 when necessary. All solutions had identical absorbance of ca. 0.3 at the 532 nm excitation wavelength. A preamplified (low impedance) Ge-photodiode cooled at 77 K (Applied Detector Corp., Model 403HS, time resolution 300 ns) and equipped with a 5 mm thick AR coated silicon metal filter with wavelength pass >1.1  $\mu$ m and an interference filter at 1.27  $\mu$ m was used to measure emission of singlet oxygen at 1270 nm in right angle geometry. The photodiode output current was fed into a digital oscilloscope. The intensities of singlet oxygen emission of reference and sample can be compared directly with each other and the calculation of the singlet oxygen emission quantum yield,  $\Phi_{\Lambda}$ , is based on the following equation

$$\Phi_{\!\Delta} = \Phi_{\!\Delta}^{\rm ref} \times I_0 / I_0^{\rm ref} \tag{4}$$

where  $I_0$  is the emission intensity extrapolated at time zero upon single exponential fitting of the emission decay with the exclusion of the initial part of the signal affected by scattered light, sensitizer fluorescence, and formation kinetics. The standard value of 0.76 was used for  $\Phi_{\Delta}^{\rm ref}$  of TPPS.<sup>40</sup>

Fluorescence Quenching and Potentiometric Titrations. Spectrofluorimetric studies were performed with on a commercial spectrophotometer. The pH-metric titrations were carried out with a commercial titration system. All titrations were performed at 25.0  $\pm$  0.1 °C. Protonation constants of ligand L were determined in a water mixture, made 0.1 M in NaNO<sub>3</sub>. In a typical experiment, 15 mL of a 5  $\times$  10<sup>-4</sup> M ligand solution were treated with an excess of a 1.0 M HNO<sub>3</sub> standard solution. Titrations were run by addition of 10  $\mu$ L aliquots of carbonate-free standard 0.1 M NaOH, recording 80–100 points for each titration. Prior to each potentiometric titration, the standard electrochemical potential (E°) of the glass electrode was determined in the water, by a titration experiment according to the Gran method.<sup>43</sup> Protonation titration data (emf vs mL of NaOH) were processed with the Hyperquad package,<sup>44</sup> to determine the equilibrium constants.

#### ASSOCIATED CONTENT

#### Supporting Information

Additional absorption and fluorescence spectra (Figure 1–6S), time-correlated single photon counting experiments (TCSPC, Figure 7S), HPLC purity data, and NMR spectra of the new NDIs. This material is available free of charge via the Internet at http://pubs.acs.org

#### AUTHOR INFORMATION

#### **Corresponding Author**

\*(M.F.) Fax: +39 0382 987323. Tel: +39 0382 987668. E-mail: mauro.freccero@unipv.it. (I.M.) E-mail: ilse.manet@isof.cnr.it. Notes

The authors declare no competing financial interest.

## ACKNOWLEDGMENTS

Financial support from MIUR, Rome (FIRB-IdeasRBI-D082ATK and PRIN 2009MFRKZ8), and University of Pavia is gratefully acknowledged. I.M. likes to thank Dr. F. Manoli for assistance in practical laboratory issues.

#### REFERENCES

(1) Brown, S. B.; Brown, E. A.; Walker, I. Lancet Oncol. 2004, 5, 497–508.

(2) Jiang, X. J.; Lo, P. C.; Tsang, Y. M.; Yeung, S. L.; Fong, W. P.; Ng, D. K. *Chem.—Eur. J.* **2010**, *16*, 4777–4783.

- (3) Lo, P. C.; Huang, J. D.; Cheng, D. Y.; Chan, E. Y.; Fong, W. P.; Ko, W. H.; Ng, D. K. *Chem.—Eur. J.* **2004**, *10*, 4831–4838.
- (4) Schweitzer, C.; Schmidt, R. Chem. Rev. 2003, 103, 1685–1757.
- (5) Stefflova, K.; Chen, J.; Zheng, G. Curr. Med. Chem. 2007, 14, 2110–2125.
- (6) Barker, C. A.; Zeng, X. S.; Bettington, S.; Batsanov, A. S.; Bryce, M. R.; Beeby, A. *Chem.—Eur. J.* **2007**, *13*, 6710–6717.
- (7) Ishii, K. Coord. Chem. Rev. 2012, 256, 1556-1568.

(8) Jiang, X. J.; Yeung, S. L.; Lo, P. C.; Fong, W. P.; Ng, D. K. J. Med. Chem. 2011, 54, 320-330.

(9) Strassert, C. A.; Otter, M.; Albuquerque, R. Q.; Hone, A.; Vida, Y.; Maier, B.; De Cola, L. *Angew. Chem., Int. Ed. Engl.* **2009**, *48*, 7928–7931.

(10) Tu, J.; Wang, T. X.; Shi, W.; Wu, G. S.; Tian, X. H.; Wang, Y. H.; Ge, D. T.; Ren, L. *Biomaterials* **2012**, *33*, 7903–7914.

(11) Wu, W.; Guo, H.; Wu, W.; Ji, S.; Zhao, J. J. Org. Chem. 2011, 76, 7056–7064.

(12) Sakai, N.; Mareda, J.; Vauthey, E.; Matile, S. Chem. Commun. 2010, 46, 4225-4237.

(13) Collie, G. W.; Promontorio, R.; Hampel, S. M.; Micco, M.; Neidle, S.; Parkinson, G. N. J. Am. Chem. Soc. **2012**, 134, 2723–2731.

(14) Cuenca, F.; Greciano, O.; Gunaratnam, M.; Haider, S.; Munnur, D.; Nanjunda, R.; Wilson, W. D.; Neidle, S. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 1668–1673.

(15) Micco, M.; Collie, G. W.; Dale, A. G.; Ohnmacht, S. A.; Pazitna, I.; Gunaratnam, M.; Reszka, A. P.; Neidle, S. *J. Med. Chem.* **2013**, *56*, 2959–2974.

(16) Di Antonio, M.; Doria, F.; Richter, S. N.; Bertipaglia, C.; Mella, M.; Sissi, C.; Palumbo, M.; Freccero, M.; Di Antonio, M.; Doria, F.; Richter, S. N.; Bertipaglia, C.; Mella, M.; Sissi, C.; Palumbo, M.; Freccero, M. J. Am. Chem. Soc. **2009**, 131, 13132–13141.

(17) Doria, F.; Nadai, M.; Folini, M.; Di Antonio, M.; Germani, L.; Percivalle, C.; Sissi, C.; Zaffaroni, N.; Alcaro, S.; Artese, A.; Richter, S. N.; Freccero, M. *Org. Biomol. Chem.* **2012**, *10*, 2798–2806.

(18) Doria, F.; Nadai, M.; Folini, M.; Scalabrin, M.; Germani, L.; Sattin, G.; Mella, M.; Palumbo, M.; Zaffaroni, N.; Fabris, D.; Freccero, M.; Richter, S. N. *Chem.—Eur. J.* **2013**, *19*, 78–81.

(19) Nadai, M.; Doria, F.; Di Antonio, M.; Sattin, G.; Germani, L.; Percivalle, C.; Palumbo, M.; Richter, S. N.; Freccero, M. *Biochimie* **2011**, 93, 1328–1340.

(20) Siddiqui-Jain, A.; Grand, C. L.; Bearss, D. J.; Hurley, L. H. Proc. Natl. Acad. Sci. U.S.A. **2002**, *99*, 11593–11598.

(21) Huppert, J. L.; Balasubramanian, S. Nucleic Acids Res. 2005, 33, 2908–2916.

(22) Huppert, J. L.; Balasubramanian, S. Nucleic Acids Res. 2007, 35, 406-413.

(23) Rodriguez, R.; Miller, K. M.; Forment, J. V.; Bradshaw, C. R.; Nikan, M.; Britton, S.; Oelschlaegel, T.; Xhemalce, B.;

Balasubramanian, S.; Jackson, S. P. *Nat. Chem. Biol.* **2012**, *8*, 301–310. (24) Doria, F.; Nadai, M.; Sattin, G.; Pasotti, L.; Richter, S. N.;

Freccero, M. Org. Biomol. Chem. **2012**, 10, 3830–3840. (25) Röger, C.; Würthner, F. J. Org. Chem. **2007**, 72, 8070–8075.

(26) Würthner, F.; Ahmed, S.; Thalacker, C.; Debaerdemaeker, T. Chem.—Eur. J. 2002, 8, 4742–4750.

(27) Zheng, K. W.; Zhang, D.; Zhang, L. X.; Hao, Y. H.; Zhou, X.; Tan, Z. J. Am. Chem. Soc. **2011**, 133, 1475–1483.

(28) Doria, F.; di Antonio, M.; Benotti, M.; Verga, D.; Freccero, M. J. Org. Chem. 2009, 74, 8616–8625.

(29) Thalacker, C.; Roger, C.; Wurthner, F. J. Org. Chem. 2006, 71, 8098-8105.

(30) Suraru, S. L.; Wurthner, F. J. Org. Chem. 2013, 78, 5227–5238. (31) Bhosale, R.; Perez-Velasco, A.; Ravikumar, V.; Kishore, R. S. K.; Kel, O.; Gomez-Casado, A.; Jonkheijm, P.; Huskens, J.; Maroni, P.; Borkovec, M.; Sawada, T.; Vauthey, E.; Sakai, N.; Matile, S. Angew. Chem., Int. Ed. 2009, 48, 6461–6464.

(32) Rogers, J. E.; Weiss, S. J.; Kelly, L. A. J. Am. Chem. Soc. 2000, 122, 427-436.

- (33) Bhosale, S.; Sisson, A. L.; Talukdar, P.; Furstenberg, A.; Banerji, N.; Vauthey, E.; Bollot, G.; Mareda, J.; Roger, C.; Wurthner, F.; Sakai,
- N.; Matile, S. Science 2006, 313, 84-86.
- (34) Guo, S.; Wu, W.; Guo, H.; Zhao, J. J. Org. Chem. 2012, 77, 3933–3943.
- (35) Ajayakumar, M. R.; Asthana, D.; Mukhopadhyay, P. Org. Lett. 2012, 14, 4822-4825.

(36) Di Antonio, M.; Doria, F.; Mella, M.; Merli, D.; Profumo, A.; Freccero, M. J. Org. Chem. 2007, 72, 8354–8360.

(37) Guha, S.; Goodson, F. S.; Corson, L. J.; Saha, S. J. Am. Chem. Soc. 2012, 134, 13679–13691.

(38) Fin, A.; Petkova, I.; Doval, D. A.; Sakai, N.; Vauthey, E.; Matile, S. Org. Biomol. Chem. **2011**, *9*, 8246–8252.

(39) Mosinger, J.; Micka, Z. J. Photochem. Photobiol. A **1997**, 107, 77–82.

(40) Wilkinson, F.; Helman, W. P.; Ross, A. B. J. Phys. Chem. Ref. Data 1993, 22, 113–262.

(41) Takizawa, S. Y.; Aboshi, R.; Murata, S. Photochem. Photobiol. Sci. 2011, 10, 895–903.

(42) Dy, J. T.; Ogawa, K.; Satake, A.; Ishizumi, A.; Kobuke, Y. Chem.—Eur. J. 2007, 13, 3491–3500.

(43) Pehrsson, L.; Ingman, F.; Johansson, A. Talanta 1976, 23, 769–780.

(44) Gans, P.; Sabatini, A.; Vacca, A. Talanta 1996, 43, 1739-1753.